



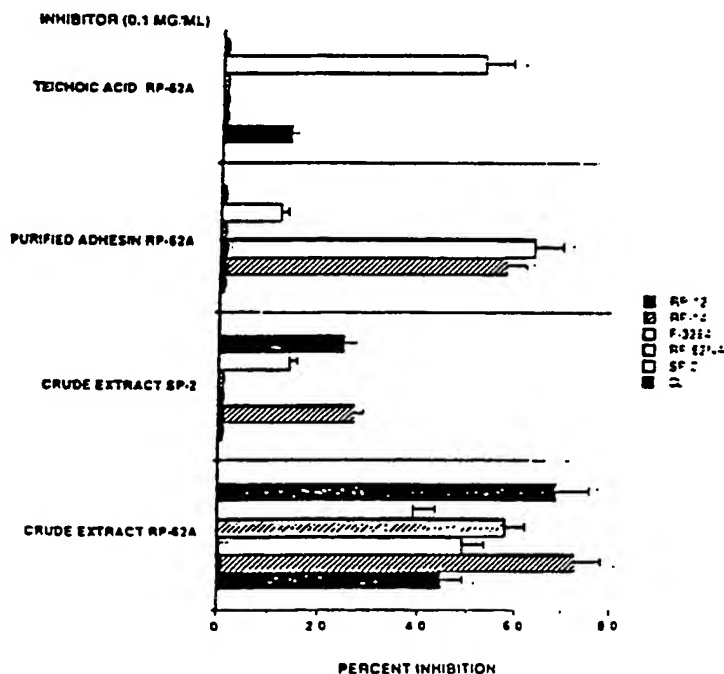
## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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(54) Title: CAPSULAR POLYSACCHARIDE ADHESIN ANTIGEN. PREPARATION. PURIFICATION AND USE

## (57) Abstract

A substantially pure capsular exopolysaccharide adhesin of coagulase-negative staphylococcal strains, and a general method to prepare such adhesins, are described. Vaccines composed of such adhesins, and uses of such adhesins to produce polyclonal and monoclonal antibodies against such adhesins, are also disclosed. The adhesins are useful in coating polymeric medical materials to prevent colonization by coagulase-negative staphylococcal strains, and as a probe in selecting desirable polymeric medical materials. Such adhesin antibodies are useful *in vivo* to prevent infection by nosocomial coagulase-negative staphylococcal strains, in assays for the detection of such bacteria, in assays for the estimation of such adhesins in complex mixtures, and as an affinity chromatography matrix.



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TITLE OF THE INVENTIONCAPSULAR POLYSACCHARIDE ADHESIN ANTIGEN,  
PREPARATION, PURIFICATION AND USEBACKGROUND OF THE INVENTIONFIELD OF THE INVENTION

The present invention relates to a substantially pure exopolysaccharide adhesin isolated from a particular strain of Staphylococcus epidermidis, to a general method capable of isolating this compound in substantially pure form, and to uses of said purified adhesin product as a vaccine for the production of antibodies effective against the binding of homologous bacterial cells to polymeric materials, and as a probe for the development of polymeric materials useful as catheters and medical prostheses.

DESCRIPTION OF THE BACKGROUND ART

Both Staphylococcus aureus (coagulase-positive) and Staphylococcus epidermidis (coagulase-negative) have a characteristic propensity for invading skin and adjacent tissues at the site of prosthetic medical devices, including intravascular catheters, cerebrospinal fluid shunts, hemodialysis shunts, vascular grafts, and extended-wear contact lenses. Within 48 to 72 hours, relatively large numbers of

staphylococci are demonstrable at the site of insertion of these foreign bodies. Archer, G.L., "Staphylococcus epidermidis: The Organism, Its Diseases, and Treatment," in Remington, J.S., et al., eds., Current Clinical Topics in Infectious Diseases, McGraw-Hill, New York, 1986, pp. 25-46; Youmans, G.P., et al., The Biologic and Clinical Basis of Infectious Diseases, Saunders, Philadelphia, 1985, pp. 618-625, 738-9. It has been demonstrated that S. epidermidis cells attach and proliferate on the inner or outer surfaces of catheters, irrespective of their composition (polyethylene, polyvinylchloride, polyvinylfluoride, or polyester based materials).

Although the virulence of coagulase-negative staphylococci clearly is enhanced in the presence of a foreign body, the microbial factor(s) that permit these normal skin commensals to become nosocomial pathogens have not been well characterized. As adherence is believed to be the critical first step in the pathogenesis of coagulase-negative staphylococcal foreign-body infections, attention has focused on surface properties of these organisms that might mediate adherence to, and colonization of, polymeric prosthetic materials.

The most promising candidate for the source of a specific staphylococcal adhesin is an extracellular material often referred to as "slime." It has been hypothesized that the slime substance may protect the S. epidermidis cells against antibiotics, as well as against natural host defense mechanisms. Youmans et al., supra; Peters, G., et al., Journal of Infectious Diseases 146:479-82 (1982).

It has been known since 1972 that coagulase-negative bacteria isolated from cerebrospinal fluid shunt infections elaborate a mucoid material that stains with alcian blue and is presumably a polysaccharide. Bayston, R., et al., Develop-

mental and Medical Child Neurology 14 (Supp. 27):25-8 (1972). The extracellular polysaccharide substance of slime-producing bacteria is a loose amorphous material composed of a range of low and high molecular weight polymers composed, in general, of neutral monosaccharides such as D-glucose, D-galactose, D-mannose, L-fucose, and L-rhamnose, and also contain amino sugars, uronic acid, and polyols such as ribitol and glycerol. Gristina, A.G., Science 237:1588-95 (1987). Glucose, galactose, phenylalanine, mannose, hexosamine, phosphorous, glycine and alanine have been found as components of the slime produced by S. epidermidis strains in clinical specimens unrelated to biomaterial infections. Ichiman, J., et al., J. Appl. Bact. 51:229 (1981). Isolates of such bacteria from sites of infections are more likely to produce slime than are random isolates from skin. Ishak, M.A., et al., Journal of Clinical Microbiology 22:1025-9 (1985). Moreover, slime-producing strains adhere well to a variety of polymeric materials. Christensen, G.D., et al., Infect. Immun. 37:318-26 (1982).

Coagulase-positive staphylococci (S. aureus) are reported to produce multiple cell surface proteins which can be released from such cells by thermal extraction and which can be shown to bind to influenza virus-infected canine kidney cells. It was considered that S. aureus produces multiple cell surface protein adhesins. Sanford, B.A., et al., Infect. Immun. 52:671-5 (1986); Proc. Soc. Exp. Biol. Med. 181:104-11 (1986).

Identification of other microbial adhesins has been reported. Pier (U.S. Patent 4,285,936, August 25, 1981; U.S. Patent 4,528,458, March 25, 1986) discloses a method for partial purification of a polysaccharide antigen from Pseudomonas aeruginosa slime. Escherichia coli fimbrial protein adhesins have been identified and partially purified by

several investigators (Orskov, I., et al., Infect. Immun. 47:191-200 (1985); Chanter, H., J. Gen. Microbiol. 125:225-243 (1983); Ferreiros, C.M., et al., Rev. espanol. de fisiolog. 39:45-50 (1983); and Moch, T., et al., Proc. Natl. Acad. Sci. 84:3462-6 (1987)).

Lectin-like glycoprotein adhesins have been identified in the Bacteroides fragilis group, and a 70 kDa adhesin has been purified by affinity chromatography (Rogemond, V., et al., Infect. Immun. 53:99-102 (1986)). Monoclonal antibody affinity chromatography was used to purify a 165 kDa surface protein of Mycoplasma pneumoniae which mediates attachment of such bacteria to target cells (Leigt, D.K., et al., J. Bacteriol. 157:678-80 (1984)), and to isolate a 150kDa adhesin protein from Streptococcus sanguis FW213 (Elder, B.L., et al., Infect. Immun. 54:421-7 (1986)). A uroepithelial cell adhesin protein of 17.5 kDa was partially purified from fimbrii of Proteus mirabilis, a frequent cause of urinary tract infection (Wray, S.K., et al., Infect. Immun. 54:43-9 (1986)).

Ludwicka (Ludwicka, A., et al., Zbl. Bakt. Hyg. A 258:256-67 (1984)) fractionated by ion-exchange chromatography a phenol-saline extract of slime from S. epidermidis and obtained four crude fractions. Both the phenol-saline extract and two of the four crude fractions inhibited the attachment of bacterial cells to polymeric material. On the basis of the presence of monosaccharides in the fractions, the reaction of the fractions with lectins, and the complete inhibition of the production of the four fractions by pretreatment of the bacteria by tunicamycin (inhibitor of glycoprotein synthesis), the authors concluded that the extracellular slime substance is a complex of glycoconjugate (i.e., glycoprotein) character.

Hogt (Hogt, A.H., et al., Infect. Immun. 51:294 (1986)) have also observed that crude extracellular products from the slime of homologous strains of S. epidermidis inhibit the

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adherence of homologous bacterial cells to polymeric materials used as catheters and prostheses. No information was provided in this report as to the chemical nature of the extracellular products.

Bacterial cells and materials derived from the surface of such cells have been used as vaccines to produce antibodies directed against homologous bacteria. Frank (Frank, R., et al., French Patent Application 85-07315, published November 21, 1986) discloses a covalent conjugate between a capsular protein adhesin (MW=74 kDa) from Streptococcus mutans and a polysaccharide from the same (serotypically) organism, and the use of said conjugate as an anti-carries vaccine. Pier (Pier, G.B., et al., U.S. patents, supra) disclose a vaccine comprising a high molecular weight mucoid exopolysaccharide from Pseudomonas aeruginosa strain 2192 slime which induces in recipient animals an immunity to said organism. Sadowski (Sadowski, P., U.S. Patent 4,443,549, April 17, 1984; U.S. Patent 4,652,498, March 24, 1984; and EP 82401506.1, published April 27, 1983) discloses ~~monoclonal~~ antibodies specific for surface adhesins of Escherichia coli and Pseudomonas aeruginosa which may be used for the therapeutic treatment of diseases induced by adhesin-bearing homologous bacteria in humans and animals. Nagy (Nagy, L.K., et al., Dev. Biol. Stand. 53:189-97 (1983)) discloses multi-adhesin vaccines for the protection of the neonatal piglet against Escherichia coli infections.

#### SUMMARY OF THE INVENTION

The inventors considered that, if a substantially pure capsular polysaccharide adhesin antigen could be isolated from the slime of strains of pathogenic S. epidermidis, a vaccine could be prepared from such an antigen that could be used to

raise polyclonal antibodies in vivo in a human or animal, or monoclonal antibodies in hybridoma cells. Reasoning that adhesin-mediated colonization is required for the onset of pathogenesis, the inventors conceived that the polyclonal or monoclonal antibodies produced against the adhesin of the invention, by preventing the adherence of adhesin-bearing pathogenic bacteria to the recipient's tissue cells or polymeric medical prostheses or catheters, represent a new means for preventing or treating diseases and infections due to S. epidermidis.

Further, the substantially pure capsular polysaccharide adhesin of the invention is useful as a probe to test new polymeric materials for medical devices.

Therefore, in a preferred embodiment, the present invention provides a substantially pure polysaccharide from extracts of S. epidermidis RP-62 strain (an isolate from a patient with catheter-related bacteremia that produces copious quantities of slime) that mediates adherence to polymeric materials and also appears to be the capsule for this organism. In another preferred embodiment, the present invention provides a method for producing a substantially pure polysaccharide adhesin from extracts of S. epidermidis strain RP-62.

In another preferred embodiment, the substantially pure polysaccharide adhesin of the invention is used as a vaccine to raise in animals antibodies against said adhesin that inhibit the attachment of adhesin-bearing bacteria to polymeric materials.

The substantially pure polysaccharide of the invention may also be used as an antigen to produce monoclonal antibodies in hybridoma cells. Such monoclonal antibodies can be administered for prophylaxis or therapeutic purposes to humans



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or animals in order to prevent or reduce infections by coagulase-negative staphylococci.

In yet another preferred embodiment, the substantially pure polysaccharide adhesin of the invention is used to screen polymeric materials for resistance to attachment by bacteria.

#### DESCRIPTION OF THE DRAWINGS

Figure 1A demonstrates the immunodiffusion pattern of crude extract (A), purified teichoic acid (B) and purified adhesin (C) against antisera raised to whole cells of S. epidermidis strain RP-62A.

Figure 1B demonstrates immunoelectrophoresis of S. epidermidis antigens. Troughs were filled with antisera to strain RP-62A whole cells. A, crude extract; B, teichoic acid, C, purified adhesin; D, mixture of teichoic acid and purified adhesin.

Figure 2 demonstrates the electrophoresis pattern of restriction enzyme digests of bacterial DNA from strains RP-62A (left-hand pattern of each pair) and RP-62NA (right-hand pattern of each pair). Lanes 1 and 12, HindIII digest of phage lambda DNA; Lanes 2 and 3, undigested DNA from RP-62A and RP-62NA; Lanes 4 and 5, EcoRI digest; Lanes 6 and 7, SauIIIA digest; Lanes 8 and 9, RsaI digest; Lanes 10 and 11, ClaI digest.

Figure 3 demonstrates the inhibition of binding of S. epidermidis strain RP-62 cells to silastic catheter tubing after incubation of the tubing in the indicated concentrations of the various bacterial antigens prior to dipping in bacterial suspension ( $10^6$  cells per ml). Significant ( $p < 0.05$ , t test) inhibition was seen only with crude extracts from strain RP-62A at concentrations of 0.12-0.50 mg/ml and with purified adhesin at concentrations of 0.06-0.50 mg/ml.

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Figure 4 demonstrates the inhibition of adherence of various strains of coagulase-negative staphylococci to silastic catheter tubing by different antigen preparations (0.1 mg/ml concentration) from S. epidermidis strain RP-62A. An asterisk indicates significant ( $p < 0.05$ , t test) inhibition.

Figure 5 shows transmission electron microscopy of various strains of coagulase-negative staphylococci following incubation with normal rabbit serum, rabbit serum raised to either whole RP-62A cells or rat antiserum raised to purified adhesin and ferritin-labeled goat antibody to rabbit or rat IgG. A) is strain RP-62 stained with normal rabbit serum (X 75,000); B) strain RP-62 stained with rabbit antisera to whole cells (X 62,000); C) strain RP-62A stained with rat antiserum to purified adhesin (X 48,000); D) strain RP-14 stained with rabbit antiserum to strain RP-62A whole cells (X 35,000); E) strain RP-14 stained with rat antiserum to purified adhesin (X 65,000); and F) strain RP-62NA stained with rabbit antiserum to strain RP-62 whole cells (X 50,000). Bar in each graph represents 200 nm.

#### DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

The invention comprises the isolation in substantially pure form of an exopolysaccharide adhesin antigen from coagulase-negative staphylococci bacteria, use of said adhesin as a vaccine to raise polyclonal and monoclonal antibodies against said adhesin, use of said adhesin to prevent adherence of said bacteria to polymeric materials, and use of said adhesin as a probe to test for polymeric materials to which said bacteria will not adhere.

### Materials and Methods

Bacterial strains. The following strains were provided by Dr. Gordon Christensen, Memphis, TN, and have been described previously (Christensen, G.D., et al., Ann. Intern. Med. 96:1-10 (1982); Infect. Immun. 37:318-26 (1982)): (a) Staphylococcus epidermidis strains RP-62A (slime-producing, highly adherent, from a patient with catheter-related sepsis), RP-62NA (a variant of RP-62A which is less adherent and produces no slime by macroscopic examination), and RP-12; (b) S. hominis strain RP-14; and (c) S. haemolyticus strain SP-2.

DNA analysis of S. epidermidis strains. Bacterial cells are lysed by the enzyme lysostaphin. The lysate is digested with RNase A (Sigma) and RNase T<sub>1</sub> (Sigma) to degrade bacterial RNA, dissolved in a detergent solution such as sodium dodecyl sulfate, and the proteins digested with proteolytic enzymes such as pronase and proteinase K (Boehringer-Mannheim). DNA is extracted from the digested cells by multiple extractions into phenol, and precipitated from the phenolic solution by the addition of ethanol at -20°C at a final concentration of 60-70% alcohol. The precipitated DNA is collected by centrifugation, washed with 70% aqueous ethanol, dried in vacuo, then digested with restriction endonucleases (EcoRI, SauIIA, RsaI and ClaI (New England Biolabs, Beverly, MA)). The restriction digest is electrophoresed on a 1% agarose gel; restriction fragments are visualized by ethidium bromide staining.

Characterization of crude extracts, purified adhesin, and teichoic acid. Samples are hydrolyzed at 100°C in 6 N HCl from 4 to 48 hours prior to analysis. Reducing carbohydrate content is detected and estimated by the phenol-sulfuric acid

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reaction (Dubois, M., et al., Anal. Chem. 28:350-6 (1956)), proteins by a positive reaction in the Bradford dye test (Bradford, M., Anal. Biochem. 72:248-54 (1976)), nucleic acids by absorbance at 254 nm against a DNA standard; phosphate by a positive reaction in the method of Chen (Chen, P.S., et al., Anal. Chem. 28:1256 (1956)); lipids by gas-liquid chromatography against fatty acid methyl esters as standards (Lee, J.C., et al., Infect. Immun., in press (1987)); and amino acids and amino sugars by an amino acid analyzer (Model 121 MB, Beckman Instruments, Inc., Fullerton, CA) using a lithium citrate system. Monosaccharides are individually identified by gas liquid chromatography of the trimethylsilyl derivatized monosaccharide methyl esters (Chambers, R.C., et al., Biochem. J. 125:1009-18 (1971)) in a Hewlett-Packard 5880 instrument using simultaneous injections of identical samples onto 25-foot capillary columns of RSL-310 (Alltech Associates, Deerfield, IL) and SP-2330 (Supelco, Delfont, PA). The injector and initial oven temperatures are 140°C, which is held for 3 minutes; a 5°C/min rise to 150°C is then performed, followed by a 30°C/min rise to 210°C, which is then held for an additional 9 minutes. The flame ionization detector is maintained at 250°C. Samples are identified by retention times compared to standards. Serologic analyses can be performed by double diffusion and immunoelectrophoresis methods (Ouchterlony, O., et al., In Immunochemistry, Vol. I, Blackwell, Oxford, 1978, Chapter 19).

Adherence assays. The adherence of coagulase-negative staphylococcal strains to polymeric (i.e., silastic) catheter tubing (French 3, Jesco International Inc., San Antonio, TX) is determined as follows. An overnight culture of bacteria in tryptic soy broth is diluted to contain  $10^6$  colony-forming units (cfu)/ml. A 3 cm length of tubing fitted with a 21

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gauge needle and sterilized with ethylene oxide gas is then dipped into the culture for 15 min at room temperature. The tubing is washed in saline by vigorously agitating the tubing, as well as repeatedly drawing saline through the tubing with a 3 ml syringe fitted to the needle. Washing is continued until wash fluids contain less than 1 cfu/100  $\mu$ l. This occurs in about 3 separate washes. After discarding a 1 cm section of the tubing, bacteria adhering to the remaining 2 cm is quantified by rolling the tubing over the surface of a tryptic soy agar plate in several directions, followed by overnight incubation at 37°C. The cfu/catheter are counted the next day. The efficiency of the transfer of bacteria from plastic tubing to the agar plate can be estimated by radio-labelling the organisms by including one  $\mu$ Ci of [ $^{14}$ C]-sodium acetate in the preliminary overnight culture medium. The number of cfu adhering to the tubing before and after rolling on the agar plate is determined by liquid scintillation counting and correlated with bacterial counts obtained by plating identical samples.

Direct adherence of the purified adhesin to catheter tubing is determined by incubating a 0.5 cm length of tubing with a 0.5 mg/ml solution of adhesin in 40 mM phosphate buffer, pH 7.2, for two hours at 37°C, washing the tubing in phosphate-buffered saline 0.05% Tween 20, and performing a standard ELISA or RIA assay on the sensitized piece of tubing (Bryan, L.E. *et al.* J. Clin. Microbiol. 18:276-82 (1983)). By the term "ELISA" is intended an enzyme-linked immunoassay. By the term "RIA" is intended a radioimmunoassay.

Inhibition of adherence of bacteria to catheter tubing by crude extracts and purified adhesin is performed by incubating the catheter tubing in solutions of these materials for two hours at 37°C, washing the coated tubing in sterile saline, placing it in bacterial cultures ( $10^6$  cfu/ml), and completing

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the adherence assay as described supra. When poorly adherent strains of S. epidermidis (e.g., strains CL and SP-2) are used in inhibition assays, the input inoculum should be increased to  $10^7$  cfu/ml, which increases the number of adhering bacteria as much as 5-fold. Inhibition of adherence is calculated as follows:

$$\% \text{ inhibition} = 100 - \frac{(100) (\text{no. of cfu adhering following adhesin treatment})}{\text{no. of cfu adhering without treatment}}$$

Inhibition of adherence by rabbit antibody to purified adhesin (see infra) is performed by incubating the bacteria with the indicated concentration of normal and immune serum for 2 hr at 4°C, washing the bacteria three times in tryptic soy broth, resuspending to  $10^6$  cfu/ml tryptic soy broth, and continuing the adherence assay as described supra. Inhibition of adherence is calculated as follows;

$$\% \text{ inhibition} = 100 - \frac{(100) (\text{no. of cfu adhering with immune serum})}{\text{no. of cfu adhering with normal serum}}$$

Inhibition data should be statistically analyzed for significance by Student's t test.

Transmission electron microscopy. Transmission electron microscopy of S. epidermidis strains is performed as previously described (Pier, GB, J. Clin. Microbiol., 24:189-96 (1986). For visualization of extracellular structures, bacterial cells are incubated with either a 1:2 dilution of rabbit antibody to whole cells or undiluted rat antibody raised to purified adhesin (see infra), or with normal serum controls. After

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three saline washes, bacteria are incubated with ferritin-conjugated antibody to either rabbit or rat IgG.

#### Preparation of Crude Bacterial Extracts

Crude extracts are prepared by incubation of cell suspensions with the enzymes lysostaphin and lysozyme. Insoluble material is removed by sequential centrifugation and filtration through a micropore filter (0.45  $\mu\text{m}$ ), the filtrate is dialyzed against water, and then lyophilized (freeze-dried in vacuo at low temperature).

#### Isolation of Adhesin

Eighteen-hour cultures of S. epidermidis strains are subjected to thermal shock (95-100°C) at about pH 5.0. The mixture is brought to neutral pH (preferably 6.8) and room temperature, then clarified by sequential centrifugation and passage through a micropore filter. The clear extract is concentrated, neutralized, and the conductivity reduced (preferably to below 10 millisiemens) by repeated ultrafiltration through a 10,000 dalton cut-off membrane and washing with water. The retained concentrate, which contains macromolecules of mass greater than 10,000 daltons, is then fractionated by ion-exchange chromatography at neutral pH (preferably about 7.0); a preferred system is DEAE Zeta-Prep 250 cartridge (LKB Instruments, Rockville, MD). Adhesin is eluted by 0.2 M NaCl at neutral pH (preferably about 7.0), as determined by an adherence assay (infra). Adhesin-containing fractions are then subjected to affinity chromatography on a Concanavalin A-Sepharose column (LKB Instruments) to remove a mannan containment that is contributed by the original bacterial tryptic soy broth growth medium and that co-purifies

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with the bacterial polysaccharide adhesin. The unbound fraction is repeatedly dialyzed against water to remove salts and small molecules, then lyophilized. After reconstitution of the adhesin-containing powder in calcium-containing buffer at an acidic pH (preferably 5.0), the solution is incubated sequentially with DNase (to remove contaminating DNA), RNase (to remove contaminating RNA), and pronase (to remove contaminating protein). The purified adhesin solution is then fractionated on a molecular sieve column in an ammonium carbonate buffer at neutral pH (preferably about 7.0). Elution is monitored by measuring  $A_{206\text{ nm}}$ ; adhesin fractions eluting with a  $K_{av}$  of 0.0-0.2 are collected and pooled. This fraction contains substantially pure capsular polysaccharide adhesin.

#### Isolation of Teichoic Acid

Teichoic acid, another component of the slime of S. epidermidis, is recovered from the DEAE Zeta Prep 250 ion-exchange column used in fractionating adhesin, in the fraction eluting with a higher concentration (0.6 M) of NaCl than eluted adhesin (0.2 M). This material is then digested with nuclease enzymes as described above, protein is denatured by heating at 100°C at an acid pH (preferably about 4.0), then chromatographed on a molecular sieve column (Sephacrose CL-4B) in ammonium carbonate buffer at neutral pH. Serologically-active fractions that elute with a  $K_{av}$  of 0.33-0.57 are pooled, dialyzed, and lyophilized.

#### Adhesin Vaccine

Polyclonal antibodies. Polyclonal antibodies to epitopic regions of the purified adhesin may be raised by a plurality



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of injections of said adhesin antigen into a host animal. In a preferred embodiment, antibodies are produced in rabbits by subcutaneous administration of 0.5 mg of antigen in complete Freund's adjuvant, followed 7 days later by intravenous injections three times weekly with 0.5 mg of antigen in saline. The thrice weekly injections are performed for 3 consecutive weeks, and blood is then drawn 5 days after the last injection. Normal (pre-immune) serum is obtained in all cases.

Polyclonal antibodies to purified adhesin may also be raised in rats given three 50  $\mu$ g injections five days apart, with blood drawn 5 days after the final injection.

Polyclonal antibodies to whole cells of S. epidermidis strains are raised in rabbits as previously described (Pier, G.B., et al., J. Infect. Dis. 147:494-503 (1983)).

Monoclonal antibodies. Monoclonal antibodies are immunoglobulins directed to specific epitopic regions on an antigen. Monoclonal antibodies against the substantially pure polysaccharide adhesin of the invention can be produced by the hybridoma technology of Köhler and Milstein (Köhler, G., Science 233:1281-6 (1986); Milstein, C., Science 231:1261-8 (1986)).

Briefly, the purified adhesin is used to once-prime or hyperimmunize animal donors of antibody-producing somatic B cells (e.g., lymphocytes). Lymph nodes and spleens of immunized animals are convenient sources. Although mouse and rat lymphocytes give a higher percentage of stable fusions with mouse myeloma lines, the use of rabbit, human and frog cells is also possible. In a preferred embodiment, hyperimmunized mouse spleen cells are used to make the fused cell hybrids.

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Specialized myeloma cell lines are available for use in hybridoma-producing fusion procedures (Köhler, G., et al., Eur. J. Immunol. 6:511-9 (1976); Schulman, M., et al., Nature 276:269-70 (1978)). Methods for generating hybrids of anti-adhesin antibody producing spleen or lymph node cells and myeloma cells usually comprise mixing somatic cells with myeloma cells in a 10:1 proportion (though the proportion can vary from 20:1 to 1:1, respectively) in the presence of an agent(s) that promotes fusion. It is preferred that the same species of animal is the source of both the somatic and myeloma cells. Fusion methods have been described by Köhler and Milstein (Köhler, G., et al., Nature 256:495-7 (1975); Eur. J. Immunol. 6:511-19 (1976)), in which Sendai virus is the fusion agent, and by Gefter (Gefter, S., et al., Somatic Cell Genet. 3:231-6 (1977)), in which polyethylene glycol is the fusion agent. In a preferred embodiment, the method of Gefter et al. is modified to include dimethylsulfoxide as an additional fusion agent.

Isolation of clones and antibody detection are carried out by standard techniques. Fusion cell hybrids are selected by culturing the cells on media that support growth of hybridomas but prevent the growth of unfused myeloma cells. (The unfused somatic cells do not maintain viability in in vitro cultures and hence do not pose a problem.) In a preferred embodiment, myeloma cells lacking hypoxanthine phosphoribosyltransferase (HPRT<sup>-</sup>) are used. These cells are selected against in a hypoxanthine/aminopterin/thymidine (HAT) medium in which hybridoma cells survive due to the HPRT<sup>+</sup> genotype of the spleen cells, but unfused myeloma cells do not. Myeloma cells with different genetic deficiencies that can be selected against in media supporting the growth of genotypically competent hybrids are also possible.

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The detection of anti-adhesin antibody-producing hybrids can be achieved by any one of several standard assays, including ELISA and RIA techniques that have been described in the literature (Kennet, R., *et al.*, eds., Monoclonal Antibodies, Hybridomas: A New Dimension in Biological Analysis, Plenum, New York, 1980, pp. 376-84; Bryan, L.E., *et al.*, J. Clin. Microbiol. 18:276-82 (1983)).

Once the desired fused cell hybrids have been selected and cloned into individual anti-adhesin antibody-producing cell lines, each cell line may be propagated in either of two standard ways: injection of the hybridoma into a histocompatible animal and recovery of the monoclonal antibodies in high concentration from the body fluids of the animal (e.g., serum or ascites fluid), or propagation in vitro in tissue culture, wherein the antibody in high concentration is recoverable from the culture medium.

#### Therapeutic Use of Anti-Adhesin Antibody

Monoclonal antibodies specific to epitopic regions on the colonization-mediating adhesin, as well as the non-specific polyclonal antibodies described above, can be used clinically for the prevention or treatment of diseases caused by pathogenic bacteria producing and bearing such adhesins. For example, polyclonal and monoclonal antibodies specific for the capsular polysaccharide adhesin of the present invention can be administered to any animal species for the prevention and/or treatment of infections due to pathogenic Staphylococcus epidermidis, e.g., those that colonize polymeric implanted medical devices and catheters. By the term "administer" is intended, for the purpose of this invention, any method of treating an animal with a substance, such as orally, intranasally, or parenterally (intravenously, intramuscularly,

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or subcutaneously). By the term "animal" is intended any living creature that is subject to staphylococcal infection, including humans, farm animals, domestic animals, or zoological garden animals. The mode of administration of these antibodies is preferably parenteral. The antibodies may be suspended or dissolved in any of several suitable liquid vehicles and delivered to the patient by any one of several parenteral means. In some instances, and particularly where human treatment is involved, purification may be desired or required pursuant to government regulations. Provided the antibody is present in a pharmacologically effective dosage, other liquid compositions are also pharmaceutically effective, including mixtures of antibodies and skim milk and/or antibodies in aqueous salt solutions of serum albumin. In humans, the antibody may be preferably administered in parenteral form, though any compatible carrier may be used. Of course, the dosage administered will be dependent upon the age, health, and weight of the recipient, kind of concurrent treatment, if any, frequency of treatment, and the nature of the effect desired. Preferably, the dosage should result in a concentration of at least about one  $\mu\text{g}$  of specific antibody per milliliter of blood.

#### Diagnostic Use of Anti-Adhesin Antibody

The adhesin-specific antibodies are also useful for medical and research purposes. For example, the antibodies can be used diagnostically to detect with great accuracy the presence of Staphylococcus epidermidis strains among a general population of bacteria. Other applications include the use of adhesin-specific monoclonal antibodies in affinity chromatography systems for the purification of Staphylococcus epider-

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midis polysaccharide adhesin or in assay systems for the quantitative estimation of such adhesin.

#### Use of Purified Adhesin as a Probe

The purified capsular polysaccharide adhesin of the invention can be used in conjunction with the adherence assays described supra as a probe in designing new polymeric materials to which coagulase-negative staphylococci bacteria will not adhere. Such new polymers would be extremely beneficial to patients in whom catheters and other medical prosthetic devices and shunts are employed and who now suffer from the nosocomial effects of such bacteria.

Having now described the invention in general term, the following specific examples will serve to illustrate more fully the nature of the present invention, without acting as a limitation upon its scope.

#### EXAMPLE I

##### Isolation of Strain PR-62A Adhesin

Staphylococcus epidermidis RP-62A was grown in 15 l of tryptic soy broth in an LSL Biolaftite fermentor with aeration (0.5 l/min), stirring (200 rpm), and maintenance of the pH at 7.2 by titration with 50% acetic acid and 5 NaOH. After 18 hr growth at 37°C, the pH was adjusted to 5.0 with 50% acetic acid and the temperature of the culture raised to 95-100°C for 1 h. After cooling, the pH was adjusted to 6.8, the culture removed from the fermentor, and bacterial cells removed by centrifugation. The supernatant was passed through a 0.5  $\mu$  filter and then concentrated to about 400 ml on a Pellicon

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ultrafiltration system (Millipore Corp., Bedford, MA) using membranes with a molecular weight cut-off of 10,000 dalton. The supernatant was then diluted with 2 l deionized water and reconcentrated to 400 ml. This step was repeated until the pH of the solution was 6.8 and the conductivity was around 4.8 millisiemens. A portion (1/4) of the solution was then applied to a DEAE Zeta-prep 250 cartridge (LKB Instruments, Rockville, MD) previously equilibrated in 0.05 M Tris buffer, pH 6.8. After loading, the cartridge was washed with 600 ml of 0.05 M Tris buffer and the eluate discarded. The adhesin was then recovered in the fraction eluting with 0.2 M NaCl in 0.05 M Tris buffer, after preliminary assays determined that this molarity of NaCl eluted material which inhibited the adherence of strain RP-62A to silastic catheter tubing (see infra). The 0.2 M NaCl eluate was pooled, dialyzed against numerous changes of deionized water, and lyophilized. The material was then resuspended in 0.1 M sodium acetate, pH 6.0, at 25 mg/ml. and chromatographed on an affinity column of Concanavalin A-Sepharose (LKB Instruments) to remove a mannan component from the tryptic soy broth medium which co-purified with the adhesin. The unbound adhesin-containing fraction was recovered, dialyzed against numerous changes of deionized water, and lyophilized. The material was then dissolved (25 mg/ml) in 0.1 M NaOH, 1.0 mM MgCl<sub>2</sub>, and 1.0 mM CaCl<sub>2</sub> pH 5.0, and digested with DNase (1 mg/ml) and RNase (3 mg/ml) for 16 hr at 37°C, after which time pronase (1.0 mg/ml) was added and an additional 4 hr digestion at 37°C carried out. This solution was then applied to a 2.6 x 90 cm column of Sepharose CL-4B (Pharmacia Fine Chemicals, Piscataway, NJ) equilibrated in 0.2 M ammonium carbonate, pH 6.8. Fractions (8 ml) were collected, and pools were made from fractions absorbing UV light at 206 nm that eluted with a K<sub>av</sub> of 0.0-0.2 (peak=0.02).

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## EXAMPLE II

Isolation of Strain PR-62A Teichoic Acid

Teichoic acid was recovered from the Zeta-prep 250 cartridge in the fraction eluting with 0.6 M NaCl. This material was digested with nuclease enzymes as described above, heated at 100°C, pH 4.0, for 1 h, then chromatographed on a 2.6 X 90 cm column of Sepharose CL-4B in 0.2 M ammonium carbonate. Serologically active fractions eluting with a  $K_{av}$  of 0.33-0.57 (peak=0.48) were pooled, dialyzed against deionized water, and lyophilized.

## EXAMPLE III

Chemical Components of Crude Extract,  
Teichoic Acid Fraction of Slime, and  
Purified Adhesin

Utilizing the methodology described above, a fraction isolated from the culture supernatant of S. epidermidis strain RP-62A that appeared to have the properties of an adhesin was analyzed. The chemical components of the crude extract, the isolated teichoic acid, and the purified adhesin are shown in Table 1.

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Table 1

Chemical Components Identified in  
Crude Extract, Teichoic Acid, and  
Purified Adhesin of Staphylococcus epidermidis  
strain RP-62A

<u>Component</u>	<u>Preparation</u>		
	<u>Crude Extract</u>	<u>Teichoic Acid</u>	<u>Purified Adhesin</u>
Reducing sugar	12*	20	54
Amino sugars	5	25	20
Uronic acids	2	<1**	10
Phosphate	11	14	<0.02**
Protein	3	2	1
Nucleic acids	7	1	1
Lipids	<0.01**	<0.01**	<0.01**
Unidentified	60	38	14
Monosaccharides (percent of total sugars)			
Glycerol		20	<0.1**
Glucose		20	<0.1**
Galactose		<0.1**	22
Glucosamine		10	15
Galactosamine		<0.1**	5

\* Percent of total weight.

\*\*Lower limit of detection.



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Crude extract contained numerous components, of which carbohydrate and phosphate were predominant. The teichoic acid fraction of slime was composed principally of phosphate, glycerol, glucose, and glucosamine. The purified adhesin was principally composed of carbohydrate with only low to non-detectable levels of protein, nucleic acids, and phosphate. No lipids were detected in the purified adhesin. The principal monosaccharides identified were galactose, glucosamine and galactosamine; glucose was absent. In addition, a complex chromatogram of monosaccharides indicated the presence of galacturonic and glucuronic acids, as well as smaller amounts of mannosamine, fucosamine, and neuraminic acid. Trace amounts of ribose and muramic acid were identified, likely due to low level contamination with RNA and peptidoglycan.

#### EXAMPLE IV

##### Serological Properties of Crude Extract, Teichoic Acid, and Purified Adhesin

Serologically, crude extract gave three precipitin lines in double diffusion when tested against a rabbit antisera raised against whole cells of strain RP-62A (Figure 1A), while teichoic acid and the purified adhesin gave single precipitin lines. By immunoelectrophoresis (Figure 1B), the crude extract had multiple precipitin lines against antisera to whole cells. In contrast, purified adhesin gave a single precipitin line which did not move in the electric field. Purified teichoic acid gave a strong precipitin line migrating towards the anodal end of the gel, as well as a weaker, more negatively charged line when high concentrations of antigen were used. A mixture of teichoic acid and purified adhesin

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resulted in two precipitin lines corresponding to the individual, purified components.

#### EXAMPLE V

##### Adherence of S. Epidermidis Strains to Polymeric Tubing

The adherence assay described supra was used to quantify the binding of strains of coagulase-negative staphylococci to silastic catheter tubing. When the inoculant size of strain RP-62A was varied from  $10^2$ - $10^9$  cfu/ml, linear binding was obtained between input inocula of  $10^3$ - $10^6$  cfu/ml. When  $10^6$  cfu/ml of radiolabeled bacteria were used in this adherence assay, and pieces of catheter tubing counted before and after being rolled over the tryptic soy agar plate, 67-75% of the counts were dislodged in three separate experiments, indicating that a majority of the adherent bacterial population was being measured by this technique.

Strains of coagulase-negative staphylococci were screened in the adherence assay at inocula of  $10^6$  cfu/ml. Three highly adherent strains of coagulase-negative staphylococci in addition to strain RP-62A (strains RP-12, RP-14, F-3284), and poorly adherent strains (Table 2).

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Table 2

Expression of Slime and Adhesin, and Adherence of  
Coagulase-Negative Staphylococci to  
Silastic Catheter Tubing

Strain	Species	Production of:		Mean No. CFU Adhering ( $\pm$ SD)
		Slime <sup>1</sup>	Adhesin	
RP-62A	<u>S. epidermidis</u>	+++	Pos <sup>2</sup>	233 $\pm$ 20
RP-12	<u>S. epidermidis</u>	+++	Neg	295 $\pm$ 40
RP-14	<u>S. hominis</u>	+	Pos	167 $\pm$ 24
F-3284	<u>S. epidermidis</u>	++	Pos	144 $\pm$ 3
RP-62NA	<u>S. epidermidis</u>	-	Pos <sup>3</sup>	68 $\pm$ 30
SP-2	<u>S. haemolyticus</u>	-	Neg	7 $\pm$ 7
CL	<u>S. haemolyticus</u>	-	Neg	19 $\pm$ 5

<sup>1</sup>Semi-quantitative measurement as described by Christensen, G.D., et al. Infect. Immun. 37:318-26 (1982).

<sup>2</sup>Presence (Pos) or absence (Neg) of adhesin determined by double immunodiffusion.

<sup>3</sup>Strain RP-62NA is only weakly positive for adhesin production.

Of the three additional highly adherent strains, two expressed an antigen giving a precipitin line of identity in double diffusion with the purified adhesin of strain RP-62A, while two of the poorly adherent strains expressed no detectable antigen. The adherence properties of strain RP-62NA were also evaluated in the adherence assay (Table 2). Strain RP-62NA adhered only about 1/3 as well as its parent strain, and a weak precipitin line corresponding to purified adhesin could be detected by immunodiffusion only if culture supernatants of strain RP-62NA were concentrated 10-fold.

Restriction enzyme digestion of total cellular DNA of strains RP-62A and RP-62NA indicated that the parent strain and its variant were closely related, as the digestion

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patterns using four different restriction enzymes were identical (Figure 2).

#### EXAMPLE VI

##### Properties of Purified Adhesin

The adhesin purified from S. epidermidis strain RP-62A was tested for its ability to inhibit adherence of the homologous strain to silastic catheter tubing. A dose-related inhibition of adherence was seen with both crude extract and purified adhesin prepared from strain RP-62A (Figure 3). Teichoic acid did not inhibit adherence of strain RP-62A, nor did the extract from the poorly adherent strain SP-2, prepared in a manner identical to that of crude extract from strain RP-62A. When these same materials (0.1 mg/ml) were tested for their ability to inhibit adherence of other strains of coagulase-negative staphylococci to silastic catheter tubing, only the two strains expressing the adhesin antigen were significantly ( $P < .05$ , t test) inhibited from adhering by purified adhesin (Figure 4). Some of the strains were inhibited from adhering to the catheter material by crude extract from strain RP-62A, and adherence of strain SP-2 was inhibited by teichoic acid from strain RP-62A.

In a similar fashion, rabbit antibodies raised to strain RP-62A-purified adhesin from strain RP-62A inhibited the adherence of this strain in a dose-related fashion at serum concentrations of  $\geq 0.25\%$ . Using a serum concentration of 1%, there was significant ( $P < .05$ , t test) inhibition of adherence of strains of coagulase-negative staphylococci expressing the adhesin antigen, while antigen-negative strains were not inhibited from adhering to silastic catheter tubing at this serum concentration (Table 3).

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Table 3

Inhibition of Adherence of  
Coagulase-Negative Staphylococci  
to Silastic Catheter Tubing by Rabbit Antibody to  
Adhesin Purified from Staphylococcus epidermidis  
Strain RP-62A

<u>Strain</u>	<u>Percent Inhibition of Adherence (<math>\pm</math>1 SD)<sup>1</sup></u>
Adhesin positive	
RP-62A	59 $\pm$ 17*
RP-14	53 $\pm$ 1*
F-3284	65 $\pm$ 14*
RP-62NA <sup>2</sup>	1 $\pm$ 13
RP-12 <sup>3</sup>	62 $\pm$ 8
Adhesin negative	
SP-2	17 $\pm$ 9
CL	0

<sup>1</sup>Serum concentration 1%.

<sup>2</sup> $p < 0.05$ , t test.

\*Produces greatly reduced amount of adhesin.

<sup>3</sup>Initial studies with RP-12 were negative. However, more recent studies have shown that RP-12 does in fact produce this adhesin. Apparently, the sera used initially failed to detect the production of adhesin from strain RP-12.

Silastic catheter tubing coated with the purified adhesin readily bound rabbit antibodies raised to whole cells and purified adhesin, while antibodies in pre-immunization sera had only a slight reaction with coated catheter tubing (Table 4).

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Table 4

Reaction of Rabbit Antibody  
to Purified Adhesin from  
Strain Staphylococcus epidermidis RP-62A with  
Silastic Catheter Tubing Catheter Coated with  
Purified Adhesin

<u>Serum</u> <u>(5% Concentration)</u>	<u>Reaction with:</u>	
	<u>Uncoated</u> <u>Catheter</u>	<u>Adhesin-Coated</u> <u>Catheter</u>
Pre-immune	0.150*	0.202
Immune to		
Whole cell	0.191	1.212
Purified adhesin	0.076	1.443

\*Mean  $A_{405}$  of triplicate wells containing indicated catheter.

## EXAMPLE VII

Transmission Electron Microscopy

Transmission electron microscopy was used to examine the appearance of bacterial cells of RP-62A, RP-62NA, RP-14, RP-12 and CL after treatment with normal rabbit or rat serum, rabbit antiserum raised to whole RP-62A cells, and rat antiserum raised to purified adhesin. Both of these antisera revealed an extracellular structure surrounding strains RP-62A and RP-14 that appeared to be a capsule (Figure 5A-D) which was not seen with normal serum (shown in the figure only for normal rabbit serum and strain RP-62A; all other strains treated with any normal serum looked identical to Figure 5A). Strain RP-62NA appeared to have only a slight amount of capsular

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material when reacted with antibodies to whole cells (Figure 5F) and purified adhesin (not shown), consistent with the serologic findings mentioned supra. Both strains RP-12 and CL lacked any detectable capsule using sera to RP-62A whole cells and purified adhesin (not shown).

The invention now being fully described, it will be apparent to one of ordinary skill in the art that many changes and modifications can be made thereto without departing from the spirit or scope of the invention as set forth herein.

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WHAT IS CLAIMED AS NEW AND DESIRED TO BE SECURED BY LETTERS  
PATENT OF THE UNITED STATES:

1. A capsular polysaccharide adhesin from coagulase-negative bacteria produced in substantially pure form.
2. The adhesin of claim 1, wherein said bacteria comprises Staphylococcus epidermidis strains.
3. The adhesin of claim 2, wherein said strain comprises strain RP-62A.
4. The adhesin of claim 2, wherein said strain comprises strain RP-62NA.
5. The adhesin of claim 2, wherein said strain comprises strain F-3284.
6. The adhesin of claim 2, wherein said strain comprises strain RP-12.
7. The adhesin of claim 1, wherein said bacteria comprises Staphylococcus hominus strains.
8. The adhesin of claim 7, wherein said strain comprises strain RP-14.
9. The adhesin of claim 1, wherein said adhesin inhibits adherence of homologous bacterial cells to polymeric material.



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10. The adhesin of claim 9, wherein said polymeric material comprises polymeric catheter and shunt tubing.

11. The adhesin of claim 9, wherein said polymeric material comprises a polymeric medical prosthetic device.

12. A process for purifying a capsular polysaccharide adhesin from coagulase-negative bacteria, comprising the following steps:

- (a) extracting adhesin from cell cultures;
- (b) chromatographically separating said adhesin extract of (a) on an ion-exchange column;
- (c) eluting adhesin from the column of (b);
- (d) chromatographically separating the adhesin-containing fraction(s) from (c) on an affinity column;
- (e) further purifying the adhesin from (d) by chromatographing on a molecular sieve column.

13. A vaccine against coagulase-negative staphylococci comprising a pharmalogically acceptable non-toxic vehicle containing the substantially pure capsular polysaccharide adhesin antigen specific to said staphylococci.

14. The vaccine of claim 13, wherein said vehicle is selected from among Freund's complete adjuvant, Freund's incomplete adjuvant, saline, serum albumin, and saponin.

15. A method of increasing immunity to infection by coagulase-negative bacteria in a member of a mammalian species comprising administering to said mammalian species a therapeutically effective amount of the vaccine of claim 13.

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16. The method of claim 15, wherein said mammalian species is a human.

17. The method of claim 15, wherein said coagulase-negative bacteria comprises a Staphylococcus epidermidis strain.

18. The method of claim 17, wherein said strain consists of strain RP-62A.

19. The method of claim 17, wherein said strain consists of strain RP-62NA.

20. The method of claim 17, wherein said strain consists of strain F-3284.

21. The method of claim 17, wherein said strain consists of strain RP-12.

22. The method of claim 17, wherein said coagulase-negative bacteria comprises a Staphylococcus hominus strain.

23. The method of claim 17, wherein said strain consists of strain RP-14.

24. A method of producing monoclonal antibodies against the capsular polysaccharide adhesin of coagulase-negative staphylococci, comprising: propagating a hybridoma formed by fusing a cell capable of producing antibodies against said adhesin with a myeloma cell and harvesting the antibodies produced by said hybridoma.

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25. Monoclonal antibody against capsular polysaccharide adhesin of coagulase-negative staphylococci.

26. A pharmaceutical composition comprising antibodies against a capsular polysaccharide adhesin of coagulase-negative staphylococci bacteria and a pharmaceutical carrier thereof.

27. The pharmaceutical composition of claim 26, wherein said antibodies are monoclonal antibodies.

28. The pharmaceutical composition of claim 26, wherein said antibodies are polyclonal antibodies.

29. The pharmaceutical composition of claim 26, wherein said bacteria comprises Staphylococcus epidermidis strain.

30. The pharmaceutical composition of claim 26, wherein said bacteria comprises Staphylococcus hominus strain.

31. A method for evaluating polymeric materials for coagulase-negative staphylococci bacteria adherence, comprising:

(a) contacting said polymeric material with said staphylococci in suspension culture;

(b) washing said polymeric material with saline to remove non-adhering bacteria;

(c) culturing adhering bacteria on the surface of a solid culture medium; and

(d) quantifying the number of bacterial colonies growing on said plate.

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32. A method of inhibiting adherence of coagulase-negative staphylococci to a polymeric material comprising contacting said polymeric material with a solution of the substantially pure capsular polysaccharide adhesin.

33. An affinity chromatography composition useful for isolating the capsular polysaccharide adhesin of a coagulase-negative staphylococcal strain from crude mixtures comprising a solid matrix to which is covalently bound a monoclonal antibody to the adhesin of claim 1.

1 / 5

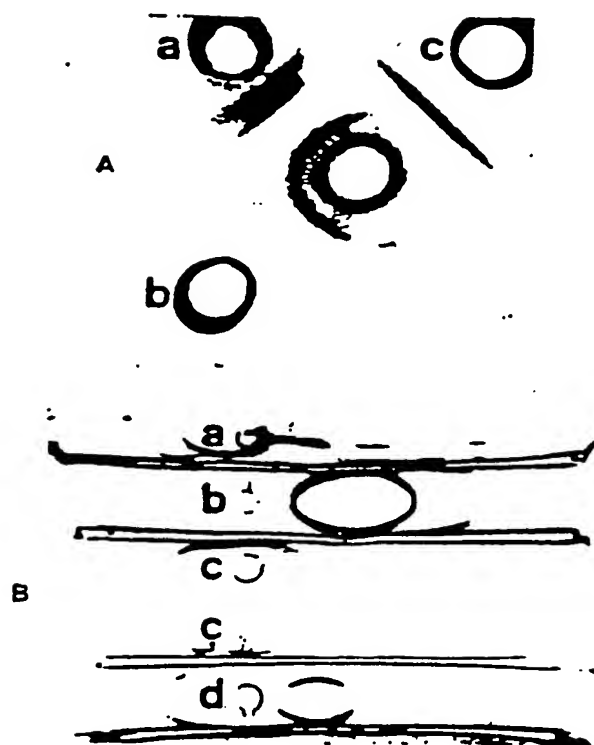


FIGURE 1

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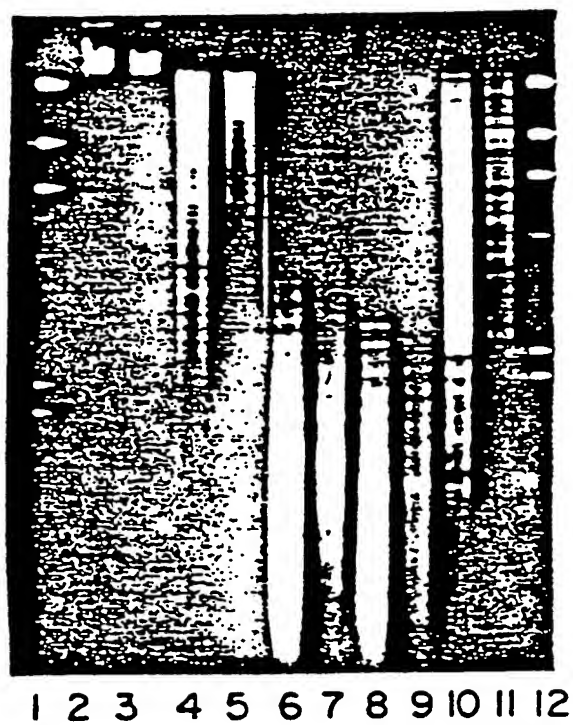


FIGURE 2

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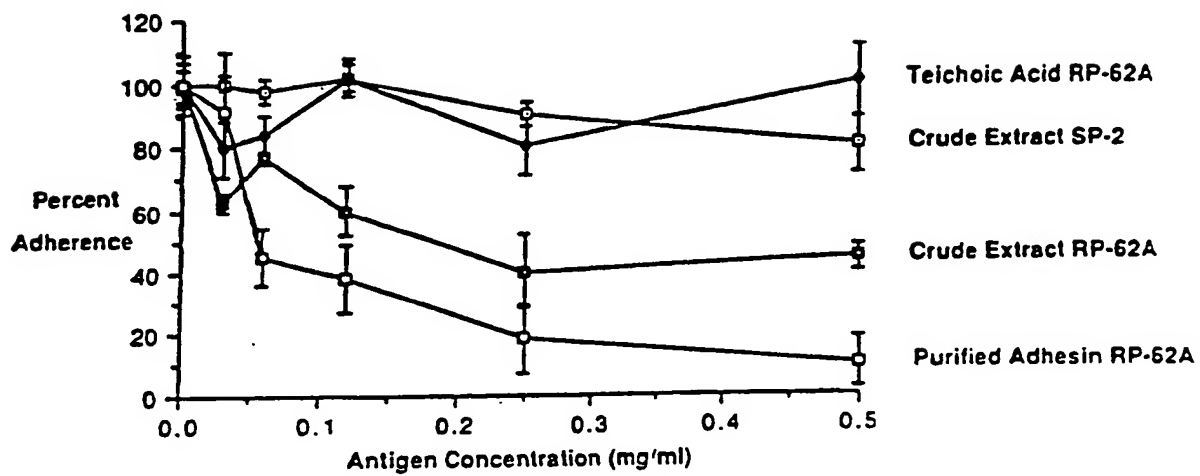


FIGURE 3

4 / 5

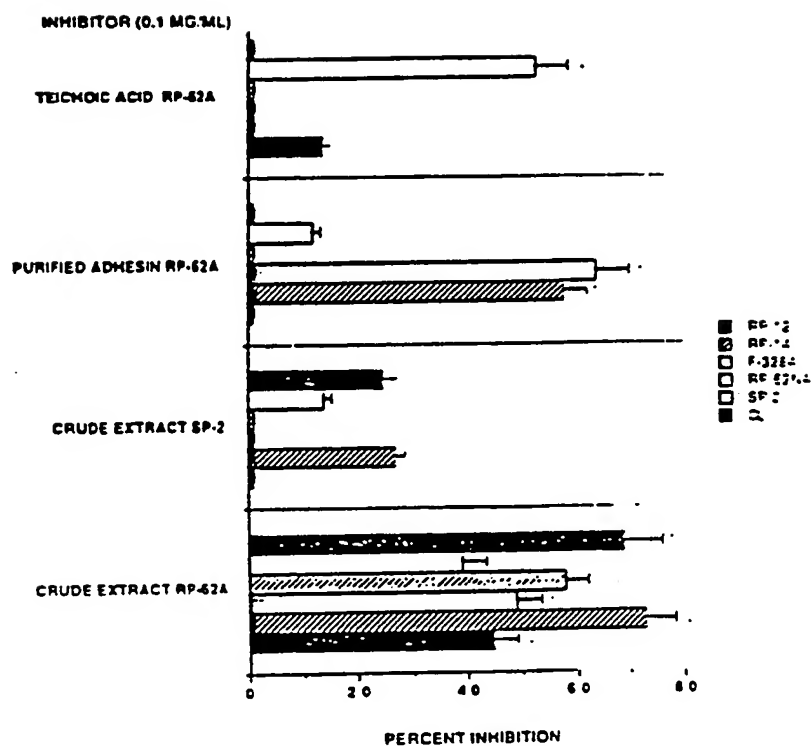


FIGURE 4



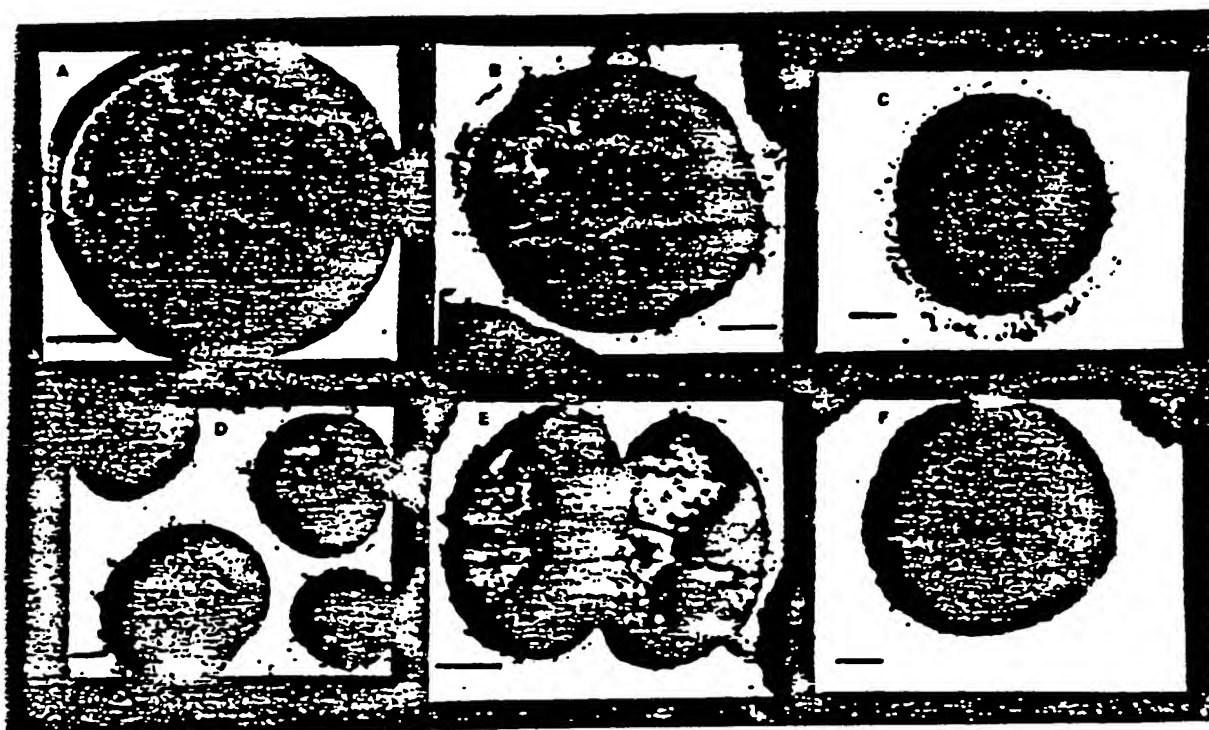


FIGURE 5

# INTERNATIONAL SEARCH REPORT

International Application No. PCT/US89/04241

## I. CLASSIFICATION OF SUBJECT MATTER (if several classification symbols apply, indicate all)

According to International Patent Classification (IPC) or to some National Classification: **IPC (4): C07K 15/04, A61K 57/00, 59/00, 59/00**

U. S. Cl.: 530/395, 402-406, 806; 536/1.1, 123, 55.1; 424/85.8, 88, 92

## II. FIELDS SEARCHED

Minimum Documentation Searched <sup>1</sup>	
Classification System	Classification Symbols
U. S.	530/395, 402-406, 806 536/1.1, 55.1, 123 424/85.8, 88, 92

Documentation Searched other than Minimum Documentation  
to the Extent that such Documents are Included in the Fields Searched<sup>2</sup>

## III. DOCUMENTS CONSIDERED TO BE RELEVANT<sup>3</sup>

Category <sup>4</sup>	Citation of Document, " with indication, where appropriate, of the relevant passages <sup>5</sup>	Relevant to Claim No. <sup>6</sup>
P, Y	US, A, 4,789,735 (Frank et al.) 06 December 1988 See entire document.	1-33
P, Y	US, A, 4,830,852 (Marburg et al.) 16 May 1989 See entire document.	1-33

### <sup>4</sup> Special categories of cited documents:

- "A" document defining the general state of the art which is not considered to be of particular relevance
- "E" earlier document but published on or after the international filing date
- "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed

- "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

- "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step

- "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

- "Z" document member of the same patent family

## IV. CERTIFICATION

Date of the Actual Completion of the International Search

21 December 1989

International Searching Authority

ISA/US

Date of Mailing of this International Search Report

10 JAN 1990

Signature of Authorized Officer

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